XCELL

QUICK GUIDE

Protocol Conditions

Cell Type Adipose-derived MSCs **Incubation** 37°C / 5% / CO_2 **Culture vessel** T75 Flask

01

Remove cultured medium from the T75 flask and wash the cells with **7 mL of DPBS**

02

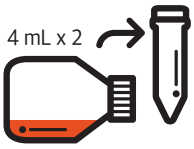
Add **2 mL of detachment reagent** into the flask and incubate cells for 3 minutes

* Using Chemically Defined products is recommended (e.g., TrypLE™ Express)

03

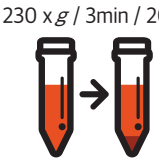
Observe under a microscope **to check for cell detachment** from the T75 flask

* Tap the flask to detach cells completely



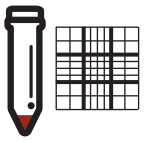
04

Collect the suspended cells in 4 mL of CellCor™ CD MSC and transfer to a tube **(Repeat once)**



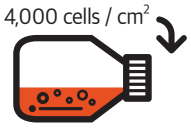
05

Centrifuge the tube at **230 x *g*, 20℃ for 3 minutes**



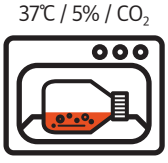
06

Remove supernatant, resuspend the cell pellet with CellCor™ CD MSC, and determine the viable cell density using a preferred method



07

Seed a T75 flask with **4,000 cells/cm²** with 15 mL of CellCor™ CD MSC



08

Incubate at 37℃ in a humidified atmosphere of 5% CO_2

09

Subculture the cells when **confluency reaches 75~85%**

* Does not change the medium until cells are 85% confluent

CAUTION

When TrypLE™ Express cannot be used, using a milder reagent instead of Trypsin is recommended (Separate neutralization phase is needed). Adhere to cell confluency percentage when culturing.

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